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Dry native proteins printed on 3D and 2D substrates by non-contact Laser-Induced-Forward Transfer (LIFT)

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Background

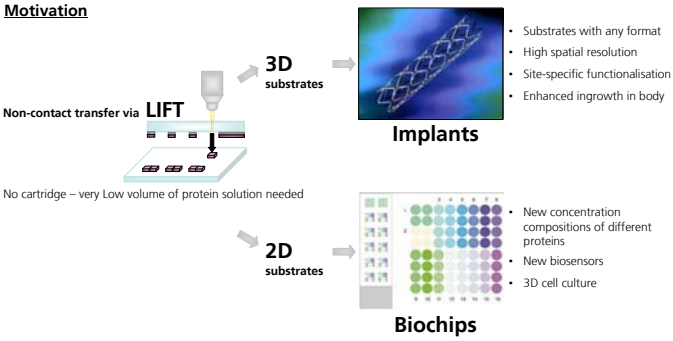
To produce nano-scale native protein trehalose patterns on 3D and 2D substrates in constant concentration by Laser-Induced-Forward Transfer (LIFT), ultrathin coatings on LIFT targets are needed [1]. The LIFT process is used as a non-contact method to transfer protein patterns on substrates at a distance of 100 µm to 300 µm. To provide selective and reproducible coatings by native pharmaceutical proteins on medical implants the LIFT process relies on homogenous protein films generated by spin-coating. In the case of 2D biochips, combined protein types with different concentrations can be arranged. Thin trehalose layers with embedded proteins such as green fluorescent protein (GFP) and the extra cellular matrix (ECM) protein laminin type1 were generated by a spin-coating technique on titanium coated targets. Robustness against long term storage is a key feature for applications, which needs multi-protein spotting (sensors; 3D cell cultures). To examine the protein stability the protein coatings were exposed to long term experiments [2,3]. Coating thickness was examined by means of ellipsometric spectroscopy on a statistically relevant number of samples [4]. During the LIFT process, the proteins are exposed to both mechanical and heat stress. Thus, subsequent tests are mandatory to grant the activity of the proteins after the transfer process. Our main focus is to examine the quality of the micro structured coatings transferred by the LIFT process using analysis by AFM and protein activity tests [3,4,5]. Activity tests are either done by application of a fluorescence scan (GFP and streptavidine with labeled biotin) or cell adhesion tests on transferred laminin type1 with fibroblasts .

Aim

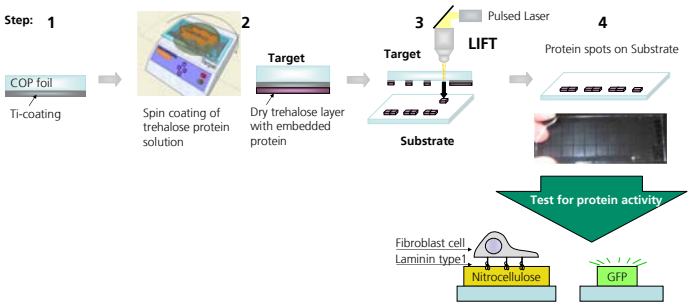
Our objective is to develop homogenous protein coatings containing native proteins of each configuration, which can be used to be patterned on 3D or 2D substrates via non-contact Laser-Induced-Forward Transfer devices.
Furthermore, we evaluate the patterned substrates applying a biosensor protein microarray, consisting of ECM protein laminin type1 for a selective fibroblast cell adsorption.

Protein printing via LIFT

Motivation

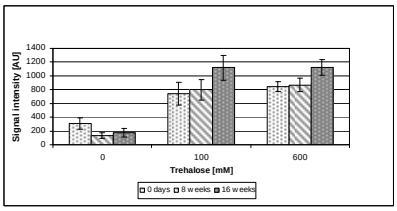


Principle workflow



Results – Robustness of proteins against drought stress

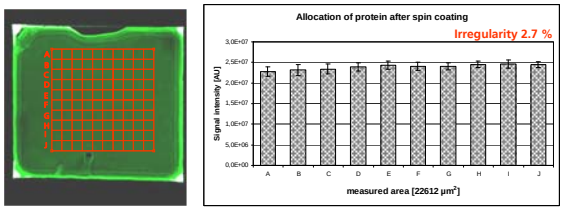
Long-term storage of streptavidine during drought stress with addition of trehalose



- Trehalose builds hydrogen bonds with proteins in drought stress and protects from denaturation.
- Trehalose = disaccharide can be found in various plants, mushrooms, hemolymph

Homogenous protein films via spin-coating

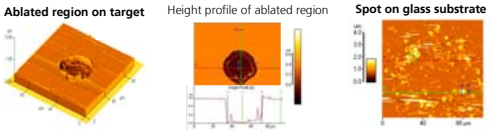
Homogenous allocation of GFP molecules within the trehalose coating



- Protein type: green fluorescent protein (GFP)
- Rotational speed: 3000 rpm
- Detection: fluorescence scan - quantitative analysis

Analysis after LIFT process

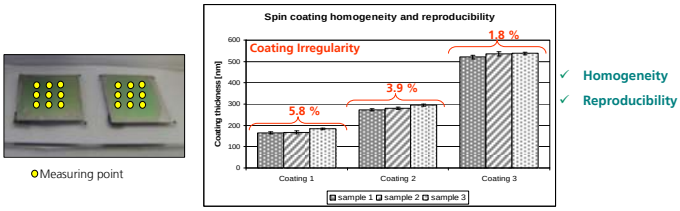
AFM analysis



Results

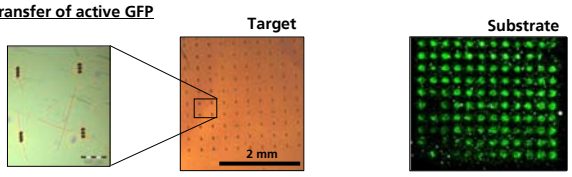
- Laser power: 6 mW
- Target
  - Homogenous protein coating
  - Ablated region:
    - Diameter 20 µm
    - Observable scope
- Height profile:
  - Sharp edges
  - Powerful ablation
  - Crater depth Ø 0.56 µm
- Opposite substrate
  - Spot well defined
  - Diameter 80 µm

Analysis of coating homogeneity and reproducibility via spectroscopic Ellipsometry



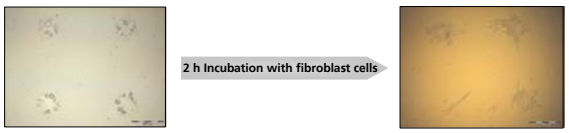
- ✓ Homogeneity
- ✓ Reproducibility

LIFT transfer of active GFP



Light microscope images — Fluorescence scan image

Selective adhesion of fibroblast cells on transferred laminin type1



LIFT transferred laminin type1 spots on glass substrate — Selective cell adhesion on the protein spots

Conclusions

- ✓ Production of dry protein trehalose coating with long-term stabilizing character on a planar target
- ✓ Creation of reproducible homogenous ultrathin protein coatings by spin coating
- ✓ Transfer of native protein patterns on the opposite substrate via LIFT

Next steps

- ➢ Decrease of satellite fragments between the spots
- ➢ Design of a Compact Disc with various proteins to create multi type protein microarrays on biochips

References

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